

Introduction

Growing awareness that the **microbial loop** plays an important role in organic matter cycling has prompted a need for new analytical tools to study microbial interactions at a molecular level. **Compound-specific stable isotope analysis of amino acids** is one such method which has shown potential for the determination of microbial reworking of organic matter [e.g. 1,2,3], however, amino acids are ubiquitous and non-specific, making them less suitable for studying the microbial food web.

Nitrogen-containing membrane lipids, on the other hand, can be highly specific: Phosphatidylethanolamine (PE) for example is common in bacteria, and much less prevalent in animals. Here we have developed a method for the determination of $\delta^{15}\text{N}$ of nitrogen-containing lipids and applied it to bacterial cultures and a microbial mat.

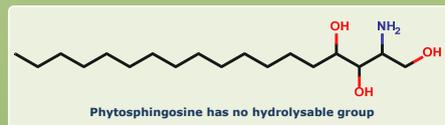
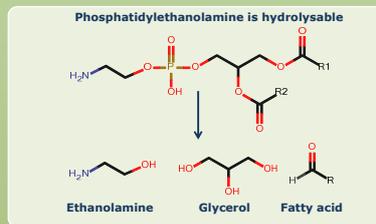
Method

Lipids were:

- Extracted using a modified Bligh & Dyer extraction [4,5]
- Acid hydrolyzed
- Prepared for GC-analysis by derivatization of polar groups using pivaloyl chloride (modified from the CSIA method for amino acid nitrogen [6, 7])
- $\delta^{15}\text{N}$ of amino acids was determined following Chikaraishi et al. [7]

Results:

- Pivaloyl chloride reacts readily with 1° and 2° amines and hydroxyl groups



The method was tested on several different N-containing lipid standards:

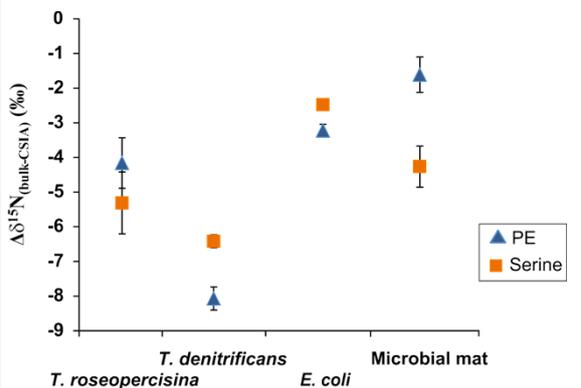
Lipid	$\Delta\delta^{15}\text{N}$ (‰) (CSIA-EA)
Hydrolysable headgroups	
Phosphatidylethanolamine (PE)	-1.2 ± 0.8
Phosphatidylserine	4.9 ± 1.5 *
n-palmitoylglycine	1.4 ± 0.3
D-erythro-sphingosine-PE [#]	2.7 ± 0.7
Cytidine phospholipid (CDP) [#]	5.1 ± 0.6 *
* Multiple products * Statistically significant difference ($\alpha=0.05$)	
No hydrolysable groups	
D-erythro-sphingosine	-0.4 ± 0.8
Phytosphingosine	0.3 ± 1.3
C-4-sphingosine	0.8 ± 0.9

- Lipids that were not hydrolyzed had small differences in $\delta^{15}\text{N}$ before and after sample workup ($\Delta\delta^{15}\text{N}$)
- Lipids that were hydrolyzed can have $\Delta\delta^{15}\text{N}$ up to 5 ‰

Conclusion: Fractionation during hydrolysis can potentially be significant

Cultures and microbial mat

The method was applied to three bacterial cultures and a microbial mat



$\delta^{15}\text{N}$ of PE and serine normalized to $\delta^{15}\text{N}$ of bulk biomass

Bacterial cultures:

- *Thiocapsa roseopersicina*: purple sulfur bacteria, photoautotroph
- *Thiobacillus denitrificans*: purple sulfur bacteria, chemoautotroph
- *Escherichia coli*: Heterotroph



Microbial mat

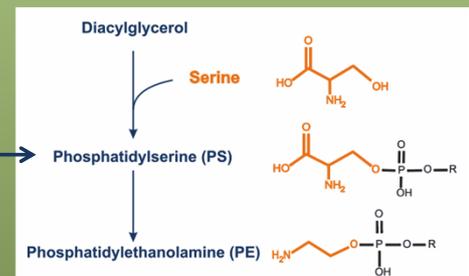
from Schiermonnikoog, Netherlands



Results:

- PE is depleted in ^{15}N compared to bulk biomass by up to 8 ‰.
- Serine is the nitrogen source to PE
- The difference in $\delta^{15}\text{N}$ between PE and serine is ~ 2 ‰:
- PE in the microbial mat is mainly influenced by heterotrophic activity
- The $\delta^{15}\text{N}$ of serine in the microbial mat has multiple sources.
- Equilibrium fractionation during hydrolysis

Biosynthetic pathway of PE



As nitrogen is not involved in the reaction, biosynthetic fractionation is not expected.

Conclusions:

- PE records the $\delta^{15}\text{N}$ signal of serine from bacteria
- Compound-specific $\delta^{15}\text{N}$ determination in lipids in combination with amino acids is a new tool to trace nitrogen through biosynthetic processes.

